DQB1*05 Product Insert Page 1 of 8

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: **09E** Lot-specific information www.olerup.com

Olerup SSP® DQB1*05

Product number: 101.211-24 – including *Taq* polymerase

101.211-24u – without *Taq* polymerase

Lot number: 09E

Expiry date: 2010-February-01

Number of tests: 24 Number of Wells per test: 6

Storage - pre-aliquoted primers: dark at -20°C

PCR Master Mix: -20°C
 Adhesive PCR seals RT
 Product Insert RT

This Product Description is only valid for Lot No. 09E.

CHANGES COMPARED TO THE PREVIOUS OLERUP SSP® DQB1*05 LOT

The DQB1*05 primer set, specificity and interpretation tables are unchanged compared to the previous *Olerup* SSP® DQB1*05 lot (Lot No. X15).

CE

February 2008

Rev. No.: 00

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: 09E Lot-specific information www.olerup.com

PRODUCT DESCRIPTION

DQB1*05 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1*0501 to DQB1*0505 alleles.

Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.

PLATE LAYOUT

Each test consists of 6 PCR reactions in an 8 well PCR plate. Wells 7 and 8 are empty.

1 2 3 4 5 6 empty empty

The 8 well cut PCR plate is marked with 'DQB1*05'.

Well No. 1 is marked with the Lot No. '09E'.

The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each 8 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Only the DQB1*05 alleles will be amplified by the DQB1*05 subtyping kit. Thus, the interpretation of DQB1*05 subtypings is not influenced by other groups of DQB1 alleles or the DQB2 and DQB3 genes.

UNIQUELY IDENTIFIED ALLELES

All the DQB1*05 alleles, i.e. **DQB1*0501 to DQB1*0505**, recognized by the HLA Nomenclature Committee in January 2008¹ will give rise to unique amplification patterns by the primers in the DQB1*05 subtyping kit.

The DQB1*05 subtyping kit cannot distinguish the DQB1*050101 and DQB1*050102 alleles and the DQB1*050301 and DQB1*050302 alleles.

¹DQB1 alleles listed on the IMGT/HLA web page 2008-January-11, release 2.20.0, www.ebi.ac.uk/imgt/hla.

RESOLUTION IN HOMO- AND HETEROZYGOTES

The 6 DQB1*05 alleles with different amplification patterns, the DQB1*050201 and DQB1*050202 alleles generate different amplification patterns, can be combined in 21 homozygous and heterozygous combinations. Eight of these genotypes do not give rise to unique amplification patterns.

```
+-++-+ 050202,0505 = 0503,0505

+-++-- 050201,050202 = 050201,0503 = 050202,050202 = 050202,0503

+-+--+ 050201,0505 = 0505,0505
```

CE

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: **09E** Lot-specific information www.olerup.com

SPECIFICITY TABLE

DQB1*05 SSP subtyping

Specificities and sizes of the PCR products of the 6 primer mixes used for DQB1*05 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified DQB1*05 ³ alleles
1	220 bp	515 bp	050101-0505
2	135 bp	430 bp	050101-050102
3	120 bp	430 bp	050201-050202, 0505
4	90 bp	515 bp	050202, 050301-050302
5	120 bp	430 bp	0504
6	185 bp	430 bp	0505

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DR low resolution SSP typings.

When the primers in a primer mix can give rise to specific PCR products of more than one length this is indicated if the size difference is 20 base pairs or more. Size differences shorter than 20 base pairs are not given. For high resolution SSP kits the respective length of the specific PCR product(s) of the alleles amplified by these primer mixes are given.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherit feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1*05 subtyping.

In addition, well number 4 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

In the presence of a specific amplification the intensity of the control band often decreases.

³For several DQB alleles only partial second exon nucleotide sequences are available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of codons 87 to 92 are conserved within allelic groups.

CE

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: **09E** Lot-specific information www.olerup.com

INTERPRETATION TABLE						
DQB1*05 SSP subtyping						
Amplification patterns of the DQB1*05 alleles						
	Well					
	1	2	3	4	5	6
Length of spec.	220	135	120	90	120	185
PCR product						
Length of int.	515	430	430	515	430	430
pos. control ¹						
5'-primer ²	26	26	30	30	30	38
	^{5'} -g gg ^{3'}	^{5'} -g gg ^{3'}	^{5'} -gA C ^{3'}	^{5'} -gA C ^{3'}	^{5'} -gA T ^{3'}	^{5'} -C gC ^{3'}
3'-primer ³	87	57	57	47	57	86
	⁵ '-g gT ³ '	5'-C AA3'	^{5'} -gCT ^{3'}	^{5'} -Cg A ^{3'}	^{5'} -gCT ^{3'}	^{5'} -A Cg ^{3'}
Well No.	1	2	3	4	5	6
DQB1 allele						
*050101-050102	1	2				
*050201	1		3			
*050202	1		3	4		
*050301-050302	1			4		
*0504	1				5	
*0505	1		3			6
DQB1 allele						
Well No.	1	2	3	4	5	6

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1*05 subtyping.

In addition, well number 4 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

²The codon, in the 2nd exon, matching the specificity-determining 3'-end of the primer is given. Codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given. Empty spaces indicate codon boundaries.

³The codon, in the 2nd exon, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given. Empty spaces indicate codon boundaries.

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: **09E** Lot-specific information www.olerup.com

DQB1*05 SSP subtyping kit Well 1 2 3 4 5	CELL LINE VALIDATION SHEET										
IHWC cell line											
IHWC cell line											
					+	1	2		_	5	6
IHWC cell line					+	-		3	4	3	U
IHWC cell line					L C	01	02	03	9	05	90
IHWC cell line					<u>İ</u>	273	273	273	273	273	200627306
IHWC cell line					odt.	790	790	790	790	790	90
1 9001 SA						20	20	20	20	20	20
2 9280 LK707		IHV	VC cell line	D	QB1						
3 9011 E4181324	1	9001	SA	*0501		+	+	•	-	•	•
### 9275 GU373		9280	LK707	*0601	*0202	-	-	-	-	-	-
5 9009 KAS011 "0502 + - + - <	3					•	-	-	-	-	-
6 9353 SM	-					-	-		-	-	-
7 9020 QBL 10201	5					+	-	+	-	-	-
8 9007 DEM		9353	SM	*0302	*0601	-	-	-	-	-	-
9 9026 YAR				_		-	-		-	-	-
10 9107 LKT3					*0502		-	-	-	-	-
11 9051 PITOUT						-	-	-	-	-	-
12 9052 DBB			_	*0401		•	-	-	-	-	-
13 9067 BTB *0402 - 14 9071 OLGA *0402 15 9075 DKB *0303 16 9037 SWEIG007 *0301 17 9008 WILJON *0602 *0603 18 9257 32367 *0602 *0202 19 9038 BM16 *0301 19 9038 BM16 *0301 20 9059 SLE005 *0604 21 9064 AMALA *0301 22 9056 KOSE *0503 *0604 + + - 23 9124 IHL *0503 *0601 + + - 24 9035 JBUSH *0301 25 9049 IBW9 *0202 26 9285 WT49 *0201 27 9191 CH1007 *0401 *0501 + + 28 9320 BEL5GB *0202 *0301 29 9050 MOU *0202 30 9021 RSH *0402 31 9019 DUCAF *0201 32 9297 HAG *0301 33 9098 MT14B *0302 34 9104 DHIF *0301 35 9302 SSTO *0305 36 9024 KT17 *0302 37 9065 HHKB *0603						-	-	-	-	-	-
14 9071 OLGA						<u> </u>	-	-	-	-	-
15						•	-	-	-	-	-
16 9037 SWEIG007 *0301 - - - - - - - - - <							-				-
17 9008 WILJON *0602 *0603 -							-				-
18 9257 32367 *0602 *0202 -				_	+0000		-		-		-
19 9038 BM16							-		-		-
20				_	*0202	-	-	-	-	-	-
21 9064 AMALA *0301						-	-	-	-	-	-
22 9056 KOSE						Ë	-	-	-	-	-
23 9124 IHL					*0604	H	-	-	-	-	-
24 9035 JBUSH *0301 - <						-	-				-
25 9049 IBW9 *0202 - <t< th=""><th></th><th></th><th></th><th></th><th>0001</th><th>Ι.</th><th>_</th><th></th><th>-</th><th></th><th>_</th></t<>					0001	Ι.	_		-		_
26 9285 WT49 *0201 - <t< th=""><th></th><th></th><th></th><th></th><th></th><th>H</th><th>-</th><th>-</th><th>-</th><th><u> </u></th><th>-</th></t<>						H	-	-	-	<u> </u>	-
27 9191 CH1007 *0401 *0501 + + -						Ė	_	_	_	_	_
28 9320 BEL5GB *0202 *0301 -					*0501	Ĥ	_	-	-	Ė	-
29 9050 MOU *0202 - <th< th=""><th></th><th></th><th></th><th></th><th></th><th></th><th>_</th><th></th><th></th><th></th><th>-</th></th<>							_				-
30 9021 RSH *0402 - 31 9019 DUCAF *0201 - 32 9297 HAG *0301 - 33 9098 MT14B *0302 - 34 9104 DHIF *0301 - 35 9302 SSTO *0305 - 36 9024 KT17 *0302 - 37 9065 HHKB *0603 - 38 9099 LZL *0301 - 39 9315 CML *0201 *0301 - 40 9134 WHONP199 *0202 *0303 - 41 9055 H0301 *0609 - 42 9066 TAB089 *0601 - 43 9076 T7526 *0303 - 44 9057 TEM *0503 + - 45 9239 SHJO *0202 - 46 9013 SCHU *0602 -					0001		-	-	-	-	-
31 9019 DUCAF *0201 - <						Ι-	-	-	-	-	-
32 9297 HAG *0301						-	-	-	-	-	-
33 9098 MT14B *0302							-		-	-	-
34 9104 DHIF *0301 - <t< th=""><th></th><th></th><th></th><th></th><th></th><th></th><th>-</th><th>-</th><th>-</th><th>-</th><th>-</th></t<>							-	-	-	-	-
35 9302 SSTO *0305 - <t< th=""><th></th><th></th><th></th><th></th><th></th><th></th><th>-</th><th>-</th><th>-</th><th>-</th><th>-</th></t<>							-	-	-	-	-
36 9024 KT17 *0302 - 37 9065 HHKB *0603 - 38 9099 LZL *0301 - 39 9315 CML *0201 *0301 - 40 9134 WHONP199 *0202 *0303 - 41 9055 H0301 *0609 - - - 42 9066 TAB089 *0601 - - - 43 9076 T7526 *0303 - - 44 9057 TEM *0503 + + - + - - 45 9239 SHJO *0202 - 46 9013 SCHU *0602 -						-	-	-	-	-	-
37 9065 HHKB *0603 - 38 9099 LZL *0301 - 39 9315 CML *0201 *0301 - 40 9134 WHONP199 *0202 *0303 - 41 9055 H0301 *0609 - - 42 9066 TAB089 *0601 - - 43 9076 T7526 *0303 - 44 9057 TEM *0503 + + - + - 45 9239 SHJO *0202 46 9013 SCHU *0602						-	-	-	-	-	-
38 9099 LZL *0301 - <td< th=""><th></th><th></th><th></th><th></th><th></th><th>-</th><th>-</th><th>-</th><th>-</th><th>-</th><th>-</th></td<>						-	-	-	-	-	-
39 9315 CML *0201 *0301 - 40 9134 WHONP199 *0202 *0303 - 41 9055 H0301 *0609 - - 42 9066 TAB089 *0601 - 43 9076 T7526 *0303 - 44 9057 TEM *0503 + + - + 45 9239 SHJO *0202 - 46 9013 SCHU *0602 -						-	-	-	-	-	-
41 9055 H0301 *0609 - 42 9066 TAB089 *0601 - 43 9076 T7526 *0303 - 44 9057 TEM *0503 + + - + 45 9239 SHJO *0202 - 46 9013 SCHU *0602 -				_	*0301		-	-	-	-	-
41 9055 H0301 *0609 - <	40					-	-	-	-	-	-
42 9066 TAB089 *0601 - 43 9076 T7526 *0303 - 44 9057 TEM *0503 + + - + + - 45 9239 SHJO *0202 - 46 9013 SCHU *0602 -	41					-	-	-	-	-	-
44 9057 TEM *0503 + + - -	42	9066	TAB089	-		-	-	-	-	-	-
45 9239 SHJO *0202 - <t< th=""><th>43</th><th></th><th></th><th></th><th></th><th>-</th><th>-</th><th>-</th><th>-</th><th>-</th><th>-</th></t<>	43					-	-	-	-	-	-
46 9013 SCHU *0602	44	9057	TEM	*0503		+	-	-	+	-	-
	45	9239	SHJO	*0202		-	-	-	-	-	-
47 9045 TUBO *0301	46	9013	SCHU	*0602		-	-	-	-	-	-
	47	9045	TUBO	*0301		-	-	-	-	-	-
48 9303 TER-ND *0501 + +	48	9303	TER-ND	*0501		+	+	-	-	-	-

DQB1*05 Product Insert Page 6 of 8

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: **09E** Lot-specific information www.olerup.com

CERTIFICATE OF ANALYSIS

Olerup SSP® DQB1*05 SSP

Product number: 101.211-24 – including *Taq* polymerase

101.211-24u – without *Taq* polymerase

Lot number: 09E

Expiry date: 2010-February-01

Number of tests: 24 Number of Wells per test: 6

Well specifications:

Well No.	Production No.
1	2006-273-01
2	2006-273-02
3	2006-273-03
4	2006-273-04
5	2006-273-05
6	2006-273-06

The specificity of each primer solution of the kit has been tested against 48 well characterized IHWC cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solution 6 were available. The specificities of the primers in this primer solution were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer.

Results: No false positive or false negative amplifications were obtained.

Date of approval: 2008-February-05

Approved by:

Quality Control, Supervisor

DQB1*05 Product Insert Page 7 of 8

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: **09E** Lot-specific information www.olerup.com

Declaration of Conformity

Product name: Olerup SSP® DQB1*05 **Product number:** 101.211-24, 101.211-24u

Lot number: 09E

Intended use: DQB1*05 resolution histocompatibility testing

Manufacturer: Olerup SSP AB

Hasselstigen 1

SE-133 33 Saltsjöbaden, Sweden

Phone: +46-8-717 88 27 **Fax:** +46-8-717 88 18

We, *Olerup* SSP AB, hereby declare that this product, to which this Declaration of Conformity relates is in conformity with the following Standard(s) and other normative document(s) ISO 9001:2000 and ISO 13485:2006, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices, Annex III, as transposed into the national laws of the Member States of the European Union.

The Technical Documentation File is maintained at *Olerup* SSP AB, Hasselstigen 1, SE-133 33 Saltsjöbaden, Sweden.

The Authorized Representative located within the Community is: *Olerup* SSP AB.

Saltsjöbaden, Sweden 2008-February-05

Olle Olerup Managing Director DQB1*05 Product Insert Page 8 of 8

101.211-24 – including *Taq* polymerase 101.211-24u – without *Taq* polymerase

Lot No.: **09E** Lot-specific information www.olerup.com

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